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<p>(54) Title: HUMAN CD45+ AND/OR FIBROBLAST + MESENCHYMAL STEM CELLS</p> <p>(57) Abstract</p> <p>Human mesenchymal stem cells having the phenotype SH3+, CD45+ can be isolated. These precursor mesenchymal stem cells are useful for treatment of patients in need of mesenchymal stem cells.</p>				

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## HUMAN CD45+ AND/OR FIBROBLAST + MESENCHYMAL STEM CELLS

**Background of the Invention**

15           This application is based on U.S. provisional application serial no. 60/087,123 filed May 29,1998.

          Mesenchymal stem cells (MSCs) are the formative pluripotential blast cells found *inter alia* in bone marrow, blood, dermis and periosteum that are capable of  
20   differentiating into more than one specific type of mesenchymal or connective tissue (*i.e.* the tissues of the body that support the specialized elements; *e.g.* adipose, osseous, stroma, cartilaginous, elastic and fibrous connective tissues) depending upon various influences from bioactive factors, such as cytokines. The potential to  
differentiate into cells such as osteoblasts and chondrocytes is retained after isolation  
25   and expansion in culture; differentiation occurs when the cells are induced *in vitro* under specific conditions or placed *in vivo* at the site of damaged tissue.

          Epitopes on the surface of the human mesenchymal stem cells (hMSCs) are reactive with certain monoclonal antibodies known as SH2, SH3 and SH4 described  
30   in U.S. Patent No. 5,486,359. These antibodies can be used as reagents to screen and capture the mesenchymal stem cell population from a heterogeneous cell population, such as exists, for example, in bone marrow.

          Hematopoietic stem cells (HSCs) are the formative pluripotential blast cells  
35   found *inter alia* in bone marrow and peripheral blood that are capable of

differentiating into any of the specific types of hematopoietic or blood cells, such as erythrocytes, lymphocytes, macrophages and megakaryocytes. The expression of a particular antigen or antigens on the cell surface or in the cytoplasm and the intensity of expression indicate the stage of maturation and lineage commitment of the hematopoietic stem cell. Human hematopoietic stem cells (hHSCs) are reactive with certain monoclonal antibodies, such as CD34, recognized as being specific for hematopoietic cells.

Thus, human hematopoietic stem cells and human mesenchymal stem cells have been readily distinguishable by their immunospecific profiles.

### Summary of the Invention

The present invention provides a population of human mesenchymal stem cells enhanced in cells that are positive for CD45 antibody markers. As hereinafter indicated, a mesenchymal stem cell is one which is capable of differentiating into more than one specific type of mesenchymal tissue cell. Applicants have provided a population of precursor mesenchymal stem cells ("pre-MSCs") which is positive for CD45. These precursor mesenchymal stem cells can differentiate into the various mesenchymal lineages, for example, the chondrocyte, adipocyte and osteoblast lineages.

In another aspect, the present invention provides a population of human mesenchymal stem cells that are CD45 positive and positive for at least one of the SH2, SH3 or SH4 markers. The mesenchymal stem cells of the present invention are preferably positive for at least the SH3 marker. In another aspect, the precursor mesenchymal stem cells are positive for the SH2 marker.

These precursor mesenchymal stem cells may be obtained using antibodies to markers of mesenchymal and hematopoietic cells. Unexpectedly, it was found that a significant number of cells positive for select markers of mesenchymal stem cells were further characterized as being CD45 positive. CD45 is a marker commonly found on leukocytes and hematopoietic cells and not on cultured mesenchymal stem cells. Although not intending to be bound by any theory, it is believed that the

population of cells of the present invention comprises a precursor cell to more mature, though not committed, mesenchymal stem cells.

The invention further provides a method for recovering an isolated population  
5 of CD45+ human mesenchymal stem cells from bone marrow or other mesenchymal  
stem cell source of an individual by (i) obtaining bone marrow tissue or other tissue  
source of mesenchymal stem cells from a donor; (ii) isolating a population of cells  
enriched in mesenchymal stem cells therefrom; and (iii) further selecting CD45+ cells  
from the population of human mesenchymal stem cells to obtain a population of  
10 mesenchymal stem cells that are enriched in CD45+ mesenchymal stem cells.

In a further aspect, the invention provides a method for recovering an isolated  
population of CD45+ human mesenchymal stem cells that are also positive for at  
least one of SH2, SH3 or SH4 markers from bone marrow or other mesenchymal  
15 stem cell source of an individual by (i) obtaining bone marrow tissue or other source  
of mesenchymal stem cell from a donor (ii) isolating a population of cells enriched in  
mesenchymal stem cells therefrom; (iii) selecting from the cell population a  
population of mesenchymal stem cells that are positive for at least one of SH2, SH3  
or SH4 markers; and (iv) further selecting CD45+ cells from the population of human  
20 mesenchymal stem cells of step (iii) to obtain a population of mesenchymal stem cells  
that are positive for at least one of either SH2, SH3 or SH4 markers and CD45+. In a  
preferred embodiment, the CD45 cell population is at least SH3 positive.

#### **Brief Description of the Drawings**

25 Figure 1 shows FACScan histograms for the expression of CD45 cell surface  
antigens on the three fractions of human mesenchymal stem cells: Fig. 1A  
prefractionation; Fig.1B SH3 selection negative fraction; Fig.1C SH3 selection  
positive fraction.

30 Figure 2 shows the results of the calcium deposition assay described in  
Example 5.

### DETAILED DESCRIPTION OF THE INVENTION

The present invention involves the isolation and enhancement of a subset population of human mesenchymal stem cells. In one embodiment, the present invention provides a population of cells having an SH3+/CD45+ phenotype and  
5 which are believed to be a precursor mesenchymal stem cell. The human mesenchymal stem cell population of the present invention is capable of differentiation into the chondrocytic, adipocytic and osteoblast cell lineages.

The mesenchymal stem cells of the present invention can be isolated from  
10 peripheral blood or bone marrow. "Isolated" as used herein signifies that the cells are placed into conditions other than their natural environment. The term "isolated" does not preclude the later use of these cells thereafter in combinations or mixtures with other cells. A method of preparing human marrow mesenchymal stem cell cultures has been described in U.S. Pat. No. 5, 486, 359. Several techniques are known to  
15 those of skill in the art for the rapid isolation of mesenchymal stem cells. Approaches to mesenchymal stem cell isolation include leucopheresis, density gradient fractionation, immunoselection and differential adhesion separation.

The cells of the present invention are maintained in culture media which can  
20 be a chemically defined serum free media or can be a "complete medium", such as Dulbecco's Modified Eagles Medium Supplemented with 10% serum (DMEM). Suitable chemically defined serum free media are described in U.S. Serial No. 08/464,599 and WO96/39487, and "complete media" are described in U.S. Patent No. 5,486,359. Chemically Defined Medium comprises a minimum essential medium  
25 such as Iscove's Modified Dulbecco's Medium (IMDM) (Gibco), supplemented with human serum albumin, human Ex Cyte lipoprotein, transferrin, insulin, vitamins, essential and non essential amino acids, sodium pyruvate, glutamine and a mitogen. These media stimulate mesenchymal stem cell growth without differentiation.

30 The mesenchymal stem cells of the present invention isolated from peripheral blood or bone marrow may further be culture-expanded. The cells may be expanded, before or after freezing thereof. The media described herein are also suitable for the culture expansion of the mesenchymal stem cells.

The isolated mesenchymal stem cells of the present invention may further be purified. In a preferred embodiment, "purified" indicates that the cell population contains less than 5% impurities, impurities being for example, cells that are not CD45+. The purified cell population can later be used in combinations or mixtures as  
5 is appropriate..

The present invention contemplates any suitable method of employing monoclonal antibodies to separate mesenchymal stem cells from other cells, e.g., recovered from bone marrow. Accordingly, included in the present invention is a  
10 method of producing a population of mesenchymal stem cells comprising the steps of providing a cell suspension of tissue containing mesenchymal stem cells; contacting the cell suspension with one or a combination of monoclonal antibodies which recognize an epitope on the mesenchymal stem cells; and separating and recovering from the cell suspension the cells bound by the monoclonal antibodies. The  
15 monoclonal antibodies may be linked to a solid-phase and utilized to capture mesenchymal stem cells from tissue samples. The bound cells may then be separated from the solid phase by known methods depending on the nature of the antibody and solid phase.

20 Monoclonal based systems appropriate for preparing the desired cell population include magnetic bead/paramagnetic particle column utilizing antibodies for either positive or negative selection; separation based on biotin or streptavidin affinity; and high speed flow cytometric sorting of immunofluorescent-stained mesenchymal stem cells mixed in a suspension of other cells. Thus, the method of  
25 the present invention includes the isolation of a population of hMSCs and enhancement using monoclonal antibodies raised against surface antigens expressed by marrow-derived hMSCs, *i.e.* SH2, SH3 or SH4. Deposits of the cell line cultures identified as SH2, SH3 and SH4 are on deposit with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852, and are assigned the  
30 ATCC accession numbers HB 10743, BH 10744 and HB 10745, respectively. These monoclonal antibodies provide effective probes which can be utilized for identifying, quantifying, and purifying mesenchymal stem cells, regardless of their source in the body.

In one embodiment, the isolation of the cell population of the present invention may comprise utilizing a combination of one or more antibodies that recognize a known marker on mesenchymal stem cells as well as an antibody which recognizes CD45. One method for such preparation of the precursor cells of the present invention is to first select a population of cells expressing a marker identifying mesenchymal stem cells, for example, SH3 or SH2 by immunomagnetic selection of a low density human bone marrow cell sample. Alternatively, it is contemplated that the initial cell selection can be based on the CD45 marker and the cell population be further characterized using the hMSC monoclonal antibodies.

10

In another embodiment, it is contemplated that a cell population can be selected based on the CD14 marker. CD14 is a membrane protein that functions as a receptor for endotoxin (lipopolysaccharide, LPS) and is expressed strongly on the surface of monocytes, but not expressed by myeloid progenitors.

15

Thus, in one aspect, in certain embodiments described herein, the mesenchymal stem cell population Population 1 (Pop 1) is identified by FACS by the relative brightness of immunofluorescent stained antibodies bound thereto as SH2 and SH3 bright/ CD45 dim/ CD14 dim. In comparison, SH2 and SH3 are present on culture expanded MSCs; CD45 is absent on culture expanded MSCs; and CD14 is absent on culture expanded MSCs.

20

In a still further aspect of the invention, a cell population can be selected based on a fibroblast cell surface marker, for example the antifibroblast antibody found on Miltenyi antifibroblast microbeads (Miltenyi catalog # 506-01).

25

It is further contemplated that the hereinabove described methods may be applied to a population of culture-expanded mesenchymal stem cells such that cells having a Pop 1 phenotype may be isolated from the population of culture-expanded mesenchymal stem cells.

30

The present invention is directed to various methods of utilizing the CD45+ human mesenchymal stem cells of the present invention for therapeutic and/or diagnostic purposes. These uses include regenerating mesenchymal tissues which



have been damaged through acute injury, abnormal genetic expression or acquired disease; treating a host having damaged mesenchymal tissue by removal of small aliquots of bone marrow, isolation of their mesenchymal stem cells and treatment of damaged tissue with the CD45+ hMSCs combined with a suitable biocompatible carrier material for delivering the MSCs to the damaged tissue sites; producing various mesenchymal tissues; detecting and evaluating growth factors or inhibitory factors relevant to MSC self-regeneration and differentiation into committed mesenchymal lineages; and developing mesenchymal cell lineages and assaying for factors with mesenchymal tissue development.

10

The hMSCs of the present invention may be used in a variety of ways. For example, the hMSCs can be employed as part of cell replacement therapy. Specifically, the hMSCs can be infused alone or added to bone marrow cells for bone marrow transplant procedures. Other applications also particularly contemplated are orthopedic, are such as augmentation of bone formation. Other applications include, for example, the treatment of osteoarthritis, osteoporosis, traumatic or pathological conditions involving any of the connective tissues, such as a bone defects, connective tissue defects, skeletal defects or cartilage defects. It is also contemplated that exogenous genetic material can be introduced into the cells while *ex vivo*, and that the cells be readministered for production of exogenous proteins *in vivo*. Genetic modification of mesenchymal stem cells is discussed more fully in U.S. Patent No. 5,591,625.

The present invention is not limited to a specific method for recovering the cells. For example, such cells may be isolated by procedures which do not use antibodies, provided that the cells are positive for CD45 and are positive for at least one of SH2, SH3 or SH4, preferably at least SH3, and are capable of differentiating into one or more than one mesenchymal cell lineage, and preferably into most, if not all, of the mesenchymal cell lineages. In a particularly preferred embodiment, the cells are also capable of self renewal. Thus, a human mesenchymal stem cell which is SH3+ and CD45+ in accordance with the invention may be recovered by techniques other than the use of SH3+ and CD45+ antibodies. Thus, the term "human mesenchymal stem cell which is SH3+ and CD45+" means a stem cell that has both

30

markers and is capable of differentiating into more than one mesenchymal stem cell lineage.

The following examples are provided to further illustrate and describe the present invention; however, the scope of the present invention is not intended to be limited thereby.

### Example 1

#### Isolation of hMSCs from Human Bone Marrow

Bone marrow aspirates were obtained from three volunteers, donors 271, 281 and 332. Density centrifugation of the bone marrow aspirates was performed using Activated Cell Therapy (ACT) Bouyant Density solution (1.0720 g/ml) in conical tubes (Dendreon, Mountain View CA) and cells were isolated from the light density fraction. Cells were washed and resuspended in Dulbecco's PBS at a concentration of  $2 \times 10^7$  cells/ml. The cells were incubated with blocking antibody (human IgG 1mg/ml in PBS azide-free) for 10 minutes at  $4^\circ \text{C}$  with rotation followed by a 30 minute incubation at  $4^\circ \text{C}$  with  $1\mu\text{g}/1 \times 10^7$  cells SH3 antibody. Cell from donors 1 and 2 were washed twice with PBS/0.5%BSA and resuspended in PBS to  $2 \times 10^7$  cells/ml. Dynal beads (washed 3 times with PBS) were added and the suspension was mixed for 30 minutes at  $4^\circ \text{C}$ . Bound cells were magnetically separated from unbound cells. Cells from donor 332 were washed 2x with Miltenyi buffer and incubated for 20 minutes at  $4^\circ \text{C}$  with mixing with rat anti-mouse IgG2 beads (1ml of microbeads per  $5 \times 10^8$  cells) (50 nm Miltenyi Biotec, Auburn, CA) and selected according to manufacturer's instructions.

Three cell populations from each donor were analyzed: the start fraction (light density unseparated cells), SH3 selected cells (cells attached to the derivatized magnetic beads) and SH3 unselected cells (cells that did not attach to the derivatized beads). The hematopoietic and mesenchymal stem cell contents of the three samples were assayed as described below.

*Cell Numbers.* Cell numbers contained in the fractions are shown in Table 1. The SH3 selected fractions contained 4.2, 4.8, and 6.2% of the starting cells. The SH3 unselected fractions yielded 88, 70, and 87% of the starting cells.

Table 1  
Cell Numbers (yields)

Fraction	Total cells ( $\times 10^6$ )	% Cell Recovery	% Viability
<b>Donor 271</b>			
Start	262		
SH3 selected	11	4.2	94.2
SH3 unselected	231	88.1	92.8
<b>Donor 281</b>			
Start	217		
SH3 selected	10.5	4.8	90.0
SH3 unselected	150	70.0	99.5
<b>Donor 332</b>			
Start	207		
SH3 Selected	12.8	6.2	
SH3 Unselected	181	87.4	

*Colony forming Unit F Assay.* The CFU-F assay measures colonies grown in complete culture media. Nucleated cells were suspended in hMSC medium to a concentration of  $2 \times 10^6$  cells in 40 mls, and were plated in 100mm tissue culture dishes at  $5 \times 10^5$  cells per plate. After 14 days cells were fixed with glutaraldehyde and stained with crystal violet. The results are shown in Table 2.

Table 2  
CFU-F Assay

Fraction	# CFU-F colonies/ $5 \times 10^4$ cells	Total CFU-F (% of total)
<b>Donor 271</b>		
Start	Not done	
SH3 selected	15	$3.3 \times 10^3$
SH3 unselected	0	0
<b>Donor 281</b>		
Start	1.3	$5.77 \times 10^3$
SH3 selected	16.7	$3.50 \times 10^3$ (61%)
SH3 unselected	0.7	$2.01 \times 10^3$ (35%)
<b>Donor 332</b>		
Start	5.7	$2.36 \times 10^4$
SH3 Selected	TNTC	
SH3 Unselected	0	0

TNTC = too numerous to count

The SH3 selected fraction showed enrichment for colonies when compared to the starting cell sample; in fact, the Miltenyi selected fraction (donor 332) had too many colonies to count. In the SH3 unselected fraction one of 3 CFU-F assays had only 0.7 colonies per 50,000 cells plated, while the remaining 2 cultures had no colony growth.

Table 3  
Hematopoietic progenitor cells Total Colonies (% Recovery)

Fraction	BFU-E	CFU-GM	CFU-GEMM
<b>Donor 271</b>			
Start	$5.61 \times 10^5$	$5.30 \times 10^5$	0
SH3 selected	0	$2.45 \times 10^2$ (0.05%)	0
SH3 unselected	$3.82 \times 10^5$ (68%)	$4.72 \times 10^5$ (89%)	0
<b>Donor 281</b>			
Start	$4.63 \times 10^5$	$2.62 \times 10^5$	0
SH3 selected	$1.16 \times 10^2$ (0.02%)	$1.16 \times 10^2$ (0.04%)	0
SH3 unselected	$3.08 \times 10^5$ (67%)	$2.56 \times 10^5$ (98%)	0
<b>Donor 332</b>			
Start	$6.07 \times 10^5$	$4.28 \times 10^5$	$8.28 \times 10^4$
SH3 Selected	0	$1.28 \times 10^3$ (0.3%)	0
SH3 Unselected	$5.61 \times 10^5$ (92%)	$4.40 \times 10^5$ (103%)	$6.03 \times 10^4$ (73%)

- 5 The SH3 unselected fractions contained 89, 98, and 103% of the starting hematopoietic CFU-GM colonies (Table 3). Only 0.04, 0.05, and 0.3% of the starting CFU-GMs were found in the SH3 selected cell fraction (Table 3).

*Mesenchymal stem cell culture.*  $3.2 \times 10^6$  cells of Donors 271 and 281 were added into 2 wells of a 6 well plate. The cells were harvested after 13 days in culture.

- 10 The results are shown in Table 4.

For sample donor 332 each well was seeded with  $0.8 \times 10^6$  cells. On Day 11, based on microscopic examination, two of four SH3 selected cell wells were harvested. All other wells were harvested after 14 days in culture. Results are shown

15 in Table 5.

- The results of the MSC cultures showed that the SH3 selected cells expanded with the same or greater efficiency than the starting cell fraction and the harvested cells had the distinctive MSC morphology and phenotype. In MSC complete culture medium after primary culture the cell yield from this SH3 unselected fraction was
- 20 low (1.3, 3.8, & 1.6%) compared to the starting cell fractions (17.5, 19.4, & 44.5%, respectively).

Table 4  
Cell numbers P0 Culture & Harvest

Fraction	Total cells harvested	% Yield
<b>Donor 271</b>		
Start	$5.6 \times 10^5$	17.5%
SH3 selected	$5.7 \times 10^5$	17.8%
SH3 unselected	$0.42 \times 10^5$	1.3%
<b>Donor 281</b>		
Start	$6.2 \times 10^5$	19.4%
SH3 selected	$6.3 \times 10^5$	19.7%
SH3 unselected	$1.2 \times 10^5$	3.8%

Table 5  
Donor 332

Fraction	Number of Cells harvested per well	Yield (% of cells seeded)	Appearance
Start	$3.56 \times 10^5$	44.5	85% confluent
SH3 Selected (Day 11)	$3.00 \times 10^5$	37.0	85% confluent
SH3 Selected (Day 14)	$5.50 \times 10^5$	68.8	100% confluent
SH3 Unselected	$1.30 \times 10^4$	1.6	no spindle shaped cells; cigar shaped or round cells

Cells from the donors 271 and 281 were continued in culture and Passage 1 cells were collected and examined by flow cytometry; the cells were MSCs by morphology and phenotype. Visual observation of these cultures during P0 showed that MSC-like colonies also contained cells with magnetic beads attached. The cells were replated in flasks or wells depending on the total cells available. The cells were harvested after 8 days in culture.

Table 6  
Cell Numbers P1 Culture & Harvest

Fraction	# Cells Plated	# Cells Harvested	Fold Increase
<b>Donor 271</b>			
Start	$4.3 \times 10^5$	$2.6 \times 10^6$	6.0
SH3 selected	$4.3 \times 10^5$	$1.8 \times 10^6$	4.2
SH3 unselected	$4.2 \times 10^4$	$0.13 \times 10^6$	3.1
<b>Donor 281</b>			
Start	$4.3 \times 10^5$	$2.3 \times 10^6$	5.3
SH3 selected	$4.3 \times 10^5$	$2.0 \times 10^6$	4.5
SH3 unselected	$1.2 \times 10^5$	$0.26 \times 10^6$	2.3

The cultured cells from the SH3 selected Day 11 harvest were analyzed. The cells were SH2+, SH3+, SH4+ and CD45-, corresponding to a mature cultured mesenchymal stem cell phenotype.

5 *Flow Cytometry Analysis.* The cell fractions of Donor 332 were analyzed using SH3 and CD45 antigen markers. Figure 1 shows the FACS histogram of the CD45 analysis.

Table 7a  
Flow Analysis Donor 332

Fraction	Total cells	% SH3 <sup>+</sup>	Total SH3 <sup>+</sup> cells	Yield (%)
Start	207 x 10 <sup>6</sup>	20.8	43.0 x 10 <sup>6</sup>	
SH3 Selected	12.8 x 10 <sup>6</sup>	98.8	12.6 x 10 <sup>6</sup>	29.4
SH3 Unselected	181 x 10 <sup>6</sup>	13.6	24.6 x 10 <sup>6</sup>	57.2

10

Table 7b

Fraction	Total cells	% SH3 <sup>+</sup> /CD45 <sup>+</sup>	Total SH3 <sup>+</sup> /CD45 <sup>+</sup> Cells	Yield (%)
Start	207 x 10 <sup>6</sup>	20.4	42.2 x 10 <sup>6</sup>	
SH3 Selected	12.8 x 10 <sup>6</sup>	98.4	12.6 x 10 <sup>6</sup>	29.8
SH3 Unselected	181 x 10 <sup>6</sup>	12.3	22.3 x 10 <sup>6</sup>	52.8

The results in Tables 7a and b of the flow analysis of the donor 332 sample demonstrated a SH3 purity of 98.8% with greater than 99.5% of these cells being  
15 CD45<sup>+</sup>. The total number of CD45- cells in the sample was 0.42%.

The results indicated that the precursor to the mesenchymal stem cell observed in culture was SH3 positive and CD45 positive and this cell can be isolated using SH3 antibody in conjunction with immunomagnetic beads or other  
20 immunoselection methods.

## Example 2

### MSC Isolation using SH2 cell selection

Biotin-anti-SH2 antibody and rat anti-mouse IgG1 magnetic microbeads  
25 were used to isolate two fractions of cells from light-density bone marrow cells: SH2 bound and SH2 unbound. These cell fractions were placed into standard MSC culture conditions to determine the MSC proliferative potential of the cell population contained in these fractions.

Anti-IgG1 microbeads were from Miltenyi Lot # NE7200. VS Column was from Miltenyi Lot # 0231). Pre-separation filters were 30 um from Miltenyi Lot # 55. Miltenyi buffer: phosphate buffered saline pH 7.2 supplemented with 0.5% BSA and 2mM EDTA. Flow staining was performed using the manufacturer's suggested instructions. Flow analysis was performed using either the FACS Calibur or the FACS Vantage. Cell viability and cell number were determined using trypan blue.

Light density cells were isolated from a human bone marrow aspirate (donor #426) using Dendreon (Seattle, WA) solution BDS72 following density centrifugation of bone marrow aspirates. Aliquots were removed as controls for flow analysis and cell culture. The remaining cells were diluted to a cell count of  $2 \times 10^7$  cells/ml using Miltenyi buffer. Cells were incubated with IgG at 40 ul per ml of cell suspension for 10 minutes at 4°C with mixing. Cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was incubated with the anti-SH2 antibody at 10 ug per  $1 \times 10^7$  cells for 30 minutes at 4°C with mixing. The cells were washed twice with cold buffer. The cell pellet was resuspended in Miltenyi buffer (80 ul of buffer per  $10^7$  cells). Anti-IgG1 microbeads were added (20 ul of beads per  $10^7$  cells). The mixture was incubated at 4°C for 15 minutes with mixing. Miltenyi buffer was added to dilute the mixture and wash the cells. The cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (0.5 ml of buffer per  $10^8$  cells). The VS column was primed following the manufacturer's instructions. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'SH2 unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'SH2 unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'SH2 bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger used to push the cells into the tube. A fresh column was primed and the cells in the 'SH2 bound' tube were added to the second column. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'SH2 unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this

effluent was added to the 'SH2 unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'SH2 bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger was used to push the cells into the tube. Cell counts and viability assays were performed. Cells from the bound and unbound fractions were stained for flow analysis. Unbound and bound cells were placed in culture. Cells were harvested after 14 days. Cells from the light-density cells and the SH2 bound fraction were stained for flow analysis.

Table 8

**SH2 Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$2.0 \times 10^8$	100
SH2 bound	$3.6 \times 10^6$	1.8
SH2 unbound	$2.3 \times 10^8$	113

Table 9

**Flow Analysis of the Starting Cell Fractions**

Fraction	% SH2+ cells
Light-density cells	23.9
SH2 bound	98.3
SH2 unbound	14.6

Table 10

**Microscopic Examination of P0 Cultures on Day 14**

Fraction	Appearance
Light-density cells	75% confluent, phase bright cells seen
SH2 Bound	90% confluent, phase bright cells seen
SH2 Unbound	Very small colonies seen

Table 11

**P0 Cell Harvests**

Fraction	Cells seeded/cm <sup>2</sup>	Cells harvested/cm <sup>2</sup>	% Yield
Light-density cells	$8.0 \times 10^4$	$1.4 \times 10^4$	17
SH2 Bound	$8.0 \times 10^4$	$1.2 \times 10^4$	15
SH2 Unbound	$8.0 \times 10^4$	$3.2 \times 10^2$	0.4



Table 12

## Flow analysis of P0 Cultures

Fraction	% MSC phenotype	% CD45+ cells
Light-density cells	99.5	0.3
SH2 Bound	98.7	0.2

5 The SH2 cell selection of light-density bone marrow cells using the Miltenyi microbead system yielded less than 2% of the light-density cells in the SH2 bound fraction. The isolation yielded a cell fraction containing cells that were 98.3% SH2+. The SH2 bound cells were 90% confluent with spindle-shaped cells after 14 days in culture under MSC standard culture conditions. The culture of the SH2 bound cells yielded a population of adherent cells that had an MSC phenotype by flow analysis and morphology. Very few adherent cells were isolated from the SH2 unbound cell fraction. These results show that SH2 is an antigen present on the MSC precursor as well as on the culture-expanded MSC.

### Example 3

### MSC Isolation of P0 MSC Cultures using CD45 Cell Selection

P0 MSC cultures have been reported to contain a median of 9% CD45+ cells (range, 0.5 to 50%), while the culture-expanded phenotype of MSCs is CD45 negative. A CD45 selection of P0 cells was performed on three donors to  
20 determine if MSCs could be cultured from the CD45 bound cell fraction

Anti-CD45 microbeads were from Miltenyi (Lot # NE5848). Large Cell Separation Column was from Miltenyi (Cat # 422-02). Miltenyi buffer: phosphate buffered saline pH 7.2 supplemented with 0.5% BSA and 2mM EDTA. Flow staining was performed using the manufacturer's suggested instructions. Flow analysis was performed using either the FACS Calibur or the FACS Vantage. Cell viability and cell number were determined using trypan blue.

P0 MSCs were derived from light density human bone marrow cells (donors # 394 (0), # 386(0) and # 381(0). Samples of  $0.5 - 5.0 \times 10^6$  cells were removed as controls for flow analysis. The remaining cells were diluted to a cell  
30 count of  $2 \times 10^7$  cells/ml using Milenyi buffer; if the count was  $< 2 \times 10^7$  cells/ml

this step was skipped. Cells were incubated with IgG at 40 ul per ml of cell suspension for 10 minutes at 4°C with mixing. Cells were centrifuged for 5 minutes at 1100 RPM. The cell pellet was resuspended in Miltenyi buffer (80 ul of buffer per  $10^7$  cells). Anti-CD45 microbeads were added (20 ul of beads per  $10^7$  cells). The mixture was incubated at 4°C for 15 minutes with mixing. Miltenyi buffer was added to dilute the mixture and wash the cells. The cells were centrifuged for 10 minutes at 1100 RPM. The cell pellet was resuspended in Miltenyi buffer (0.5 ml of buffer per  $10^8$  cells). The Large Cell Separation Column was primed following the manufacturer's instructions. The cell/microbead suspension was added to the column and the suspension drained through the column. The effluent was collected as the 'CD45 unbound' fraction. The column was rinsed three times with 0.5 ml of Miltenyi buffer and this effluent was added to the 'CD45 unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'CD45 bound'. One ml of Miltenyi buffer was added to the column and a syringe plunger used to push the cells into the tube. A fresh large cell column was primed following the manufacturer's instructions and the cells in the 'CD45 bound' tube were added to the second column. The cell/microbead suspension was added to the column and the suspension drained through the column. The effluent was collected as the 'CD45 unbound' fraction. The column was rinsed three times with 0.5 ml of Miltenyi buffer and this effluent was added to the 'CD45 unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'CD45 bound'. One ml of Miltenyi buffer was added to the column and a syringe plunger used to push the cells into the tube. Cell counts and viability assays were performed. Cells from the bound and unbound fractions (# 394) and the unbound fraction (# 386 & # 381) were stained for flow analysis. The cells were placed in culture. Cells were harvested and passaged until an adequate number of cells was available for flow analysis.

Table 13

**Donor # 381: P0 CD45 Cell selection yields**

<b>Fraction</b>	<b>Number of cells</b>	<b>% Yield</b>
P0 cells 381	$8.7 \times 10^6$	100
CD45 bound	$1.4 \times 10^5$	1.6
CD45 unbound	$4.8 \times 10^6$	55

Table 14

**Donor # 386: P0 CD45 Cell selection yields**

<b>Fraction</b>	<b>Number of cells</b>	<b>% Yield</b>
P0 cells 386	$1.2 \times 10^7$	100
CD45 bound	$2.2 \times 10^5$	1.9
CD45 unbound	$8.4 \times 10^6$	70

Table 15

**Donor # 394: P0 CD45 Cell selection yields**

<b>Fraction</b>	<b>Number of cells</b>	<b>% Yield</b>
P0 cells 394	$1.2 \times 10^8$	100
CD45 bound	$1.8 \times 10^6$	1.5
CD45 unbound	$1.1 \times 10^8$	92

Table 16

**Flow Analysis of the Starting Cell Fractions**

<b>Fraction</b>	<b>% CD45+ cells</b>
P0 cells 381	4.7
CD45 unbound 381	1.5
P0 cells 386	1.9
CD45 unbound 386	3.5
P0 cells 394	2.0
CD45 unbound 394	1.4

Table 17

**Donor 381(1) Cell Harvests**

<b>Fraction</b>	<b>Cells/cm<sup>2</sup> seeded</b>	<b>Cells/cm<sup>2</sup> harvested</b>	<b>Days in culture</b>	<b>Fold Expansion</b>
CD45 bound	$3.4 \times 10^4$	$5.8 \times 10^4$	12	1.7
CD45 unbound	$5.4 \times 10^4$	$8.3 \times 10^3$	7	1.5

Table 18

**Donor 381(2) CD45 bound Cell Harvest**

<b>Fraction</b>	<b>Cells/cm<sup>2</sup> seeded</b>	<b>Cells/cm<sup>2</sup> harvested</b>	<b>Days in culture</b>	<b>Fold Expansion</b>
CD45 bound	$2.9 \times 10^3$	$3.7 \times 10^4$	3	12.8

Table 19

**Donor 386(1) Cell Harvests**

<b>Fraction</b>	<b>Cells/cm<sup>2</sup> seeded</b>	<b>Cells/cm<sup>2</sup> harvested</b>	<b>Days in culture</b>	<b>Fold Expansion</b>
CD45 bound	$4.4 \times 10^4$	$7.5 \times 10^4$	12	1.7
CD45 unbound	$5.4 \times 10^3$	$1.6 \times 10^4$	7	3.0

5

Table 20

**Donor 394(1) Cell Harvests**

<b>Fraction</b>	<b>Cells/cm<sup>2</sup> seeded</b>	<b>Cells/cm<sup>2</sup> harvested</b>	<b>Days in culture</b>	<b>Fold Expansion</b>
CD45 bound	$5.3 \times 10^3$	$1.3 \times 10^4$	11	2.5
CD45 unbound	$5.3 \times 10^3$	$3.1 \times 10^4$	11	5.0

Table 21

**Flow Analysis of Cells Cultured from CD45 Selected P0 MSCs**

<b>Fraction</b>	<b>% MSC phenotype</b>	<b>% CD45+</b>
381(2) from CD45 bound	99.2	0.7
386(1) from CD45 bound	99.0	0.8
394(1) from CD45 bound	98.5	1.4

10 The CD45 selection of P0 MSCs yielded a cell population which was < 2% of the starting P0 population. Cultured CD45 bound cells isolated from the P0 MSCs yielded MSCs as defined by flow cytometry and morphology. The CD45 unbound cell fraction isolated from the P0 MSCs also yielded MSCs as defined by flow cytometry and morphology. The phenotype of the MSC precursor appeared to be

15 CD45 dim.

**Example 4****MSC Isolation using CD45 Selection**

20 Light-density cells isolated from bone marrow were selected using directly conjugated anti-CD45 Miltenyi microbeads (Miltenyi Lot # NE5848) following the manufacturer's instructions. VS Column was from Miltenyi (Lot # 0231). Pre-separation filters 30 um were from Miltenyi (Lot 55). Miltenyi buffer: phosphate

buffered saline pH 7.2 supplemented with 0.5% BSA and 2mM EDTA. Flow staining was performed using the manufacturer's suggested instructions. Flow analysis was performed using either the FACS Calibur or the FACS Vantage. Cell viability and cell number were determined using trypan blue.

5        Light density cells were isolated from human bone marrow aspirate (donor # 358) using Dendreon solution BDS72 following density centrifugation of the bone marrow aspirates. Aliquots were removed as controls for flow analysis and cell culture. The remaining cells were diluted to a cell count of  $2 \times 10^7$  cells/ml using Miltenyi buffer. Cells were incubated with IgG at 40 ul per ml of cell  
10 suspension for 10 minutes at 4°C with mixing. Cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (80 ul of buffer per  $10^7$  cells). Anti-CD45 microbeads were added (20 ul of beads per  $10^7$  cells). The mixture was incubated at 4°C for 20 minutes with mixing. Miltenyi buffer was added to dilute the mixture and wash the cells. The cells were  
15 centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (0.5 ml of buffer per  $10^8$  cells). The VS column was primed following the manufacturer's instructions. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected  
20 as the 'CD45 unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'CD45 unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'CD45 bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger was used to push the cells into the tube. A fresh column was primed and the cells  
25 in the 'CD45 bound' tube were added to the second column. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'CD45 unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'CD45  
30 unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'CD45 bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger was used to push the cells into the tube. Cell counts and viability assays were performed. Cells from the bound and unbound fractions were

stained for flow analysis. Cells were placed in culture. Cells were harvested and counted.

Table 22  
CD45 Cell selection yields

Fraction	Number of cells	% Yield
Light-density cells	$3.93 \times 10^8$	100
CD45 bound	$1.84 \times 10^8$	47
CD45 unbound	$1.37 \times 10^8$	35

Table 23

Flow analysis of the Starting Cell Fractions

Fraction	% CD45+ cells
Light-density cells	78.3
CD45 bound	99.0 *
CD45 unbound	51.5

\* the staining intensity is significantly enhanced in the CD45 bound cell fraction

Table 24

Microscopic Examination of P0 Cultures on Day 14

Fraction	Appearance
Light-density cells	100% confluent, many phase bright cells
CD45 bound	80% confluent, some phase bright cells
CD45 unbound	95% confluent, some phase bright cells

Table 25

P0 Cell Harvests

Fraction	Cells/cm <sup>2</sup> seeded	Cells/cm <sup>2</sup> harvested	% Yield
Light-density cells	$1.6 \times 10^5$	$5.4 \times 10^4$	33
CD45 bound	$1.6 \times 10^5$	$1.9 \times 10^4$	12
CD45 unbound	$1.6 \times 10^5$	$4.6 \times 10^4$	29

Forty-seven percent of the cells were present in the CD45 bound fraction. This CD45 bound cell population was 99% CD45 positive by flow analysis and was capable of yielding MSCs at P0 as defined by morphology. The CD45

unbound cell fraction was 35% of the total cells selected and was 51.5% CD45 positive. The fluorescence intensity of the CD45 unbound cells was much lower than that of the CD45 bound cells. This cell fraction was also capable of yielding MSCs in P0 culture as defined by morphology. This experiment provides further  
5 evidence that the MSC precursor is CD45 positive with a dim staining intensity.

### Example 5

#### MSC Isolation using Fibroblast Cell Selection

Anti-fibroblast microbeads were developed for the separation of cells based  
10 on the expression of a fibroblast-specific antigen. Since cultured MSCs have a fibroblastic morphology, the anti-fibroblast microbeads were used to select a fraction of cells from light-density bone marrow cells. Cells that were bound and those that were not bound were placed in standard MSC culture conditions and observed.

15 Anti-fibroblast microbeads were from Miltenyi (Lot # NE630). VS Column was from Miltenyi (Lot # 0231). Pre-separation filters 30 um were from Miltenyi (Lot # 55). Miltenyi buffer: phosphate buffered saline pH 7.2 supplemented with 0.5% BSA and 2mM EDTA. Flow staining was performed following the manufacturer's suggested instructions. Flow analysis was performed using either  
20 the FACS Calibur or the FACS Vantage. Cell viability and cell number were determined using trypan blue.

Assays were performed as follows to measure the osteogenic and adipogenic potential of the cells.

*Adipogenesis assay.* Cells were plated in a 6-well dish ( $2 \times 10^5$  cells/well)  
25 in hMSC media. Confluent MSCs were pulse induced with high glucose media containing dexamethasone, insulin, 3-isobutyl-1-methyl-xanthine and indomethacin. At the end of the culture period the plates were fixed with 10% formalin, stained with Oil Red "O" and counterstained with hematoxylin. The formation of lipid vacuoles which stain red were observed and semi-quantitated by  
30 percent of well surface area.

*Osteogenic calcium deposition assay.* Cells were plated in 6-well dishes ( $3 \times 10^4$  cells/well). Wells labeled as "OS" were fed with hMSC media containing supplements of Ascorbic acid-2-phosphate, dexamethasone and  $\beta$ -

glycerophosphate. Wells labeled as "Control" were fed with standard hMSC media. Media changes were performed twice weekly for 14 to 16 days. The increased calcium deposition was measured through semi-quantitative colorimetric assays.

5

Light density cells were isolated from human bone marrow aspirates (donors #373, # 386 & # 421) using Dendreon solution BDS72 following density centrifugation of the bone marrow aspirates. Aliquots were removed as controls for cell culture. The remaining cells were diluted to a cell count of  $2 \times 10^7$  cells/ml using Miltenyi buffer. Cells were incubated with IgG at 40 ul per ml of cell suspension for 10 minutes at 4°C with mixing. Cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (80 ul of buffer per  $10^7$  cells). Anti-fibroblast microbeads were added (20 ul of beads per  $10^7$  cells). The mixture was incubated at room temperature for 30 minutes with mixing. Miltenyi buffer was added to dilute the mixture and wash the cells. The cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (1 ml of buffer per  $10^8$  cells). The VS column was primed following the manufacturer's instructions. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'fibroblast unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'fibroblast unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'fibroblast bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger was used to push the cells into the tube. A fresh column was primed and the cells in the 'fibroblast bound' tube were added to the second column. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'fibroblast unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'fibroblast unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'fibroblast bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger



was used to push the cells into the tube. Cell counts and viability assays were performed. Cells from donors 373 and 386 were placed in culture. Cells were harvested and passaged. P0 cells from the light density control and the fibroblast bound cultures from donor 373 were stained for flow analysis. P0 cells from donor 386 were placed into the *in vitro* osteogenic differentiation assay and the *in vitro* adipogenic assay. Cultures from the adipogenic assays showed significant adipogenesis of the MSCs.

Table 26

**Donor 373 Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$6.0 \times 10^7$	100
Fibroblast bound	$5.6 \times 10^6$	9
Fibroblast unbound	$4.2 \times 10^7$	70

Table 27

**Donor 386 Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$6.0 \times 10^8$	100
Fibroblast bound	$2.1 \times 10^7$	3.5
Fibroblast unbound	$4.3 \times 10^8$	72

Table 28

**Donor 421 Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$4.4 \times 10^8$	100
Fibroblast bound	$2.6 \times 10^7$	5.8
Fibroblast unbound	$4.2 \times 10^8$	94

Table 29

**Donor 373 Microscopic Examination of P0 Cultures on Day 14**

Fraction	Appearance
Light-density cells	80% confluent, phase bright cells seen
Fibroblast bound	50% confluent, phase bright cells seen
Fibroblast unbound	Round cells, debris

Table 30

**Donor 386 Microscopic Examination of P0 Cultures on Day 14**

Fraction	Appearance
Light-density cells	100% confluent, phase bright cells seen
Fibroblast bound	100% confluent, phase bright cells seen
Fibroblast unbound	Round floating cells, debris

Table 31

**Donor 373 P0 Cell Harvests**

Fraction	Cells seeded/cm <sup>2</sup>	Cells harvested/cm <sup>2</sup>	% Yield
Light-density cells	8.1 x 10 <sup>4</sup>	1.2 x 10 <sup>4</sup>	15
Fibroblast bound	8.0 x 10 <sup>4</sup>	5.0 x 10 <sup>3</sup>	10
Fibroblast unbound	8.1 x 10 <sup>4</sup>	3.2 x 10 <sup>1</sup>	0.04

5

Table 32

**Donor 386 P0 Cell Harvests**

Fraction	Cells seeded/cm <sup>2</sup>	Cells harvested/cm <sup>2</sup>	% Yield
Light-density cells	1.6 x 10 <sup>5</sup>	1.6 x 10 <sup>4</sup>	10
Fibroblast bound	1.6 x 10 <sup>5</sup>	1.8 x 10 <sup>4</sup>	11
Fibroblast unbound	1.6 x 10 <sup>5</sup>	3.9 x 10 <sup>1</sup>	0.02

Table 33

**Donor 373 Flow analysis of P0 Cultures**

Donor/Fraction	% MSC phenotype	% CD45+ cells
373(0) light-density cells	89.3	10.3
373(0) fibroblast bound cells	85.9	13.8

10

The results of the calcium assay for Donor 386(1), Light-density cell and fibroblast bound cell fractions are shown in Table 39 and Figure 2.

Table 34

Sample ID	Condition	Dilution Factor	Spectrophotometer OD Readings at 575nm			Calcium (ug/well)			Mean	S.D.
			1	2	3	1	2	3		
Light density	Control	20	0.0000	0.0000	0.0000	0.0	0.0	0.0	0.0	0.0
Light density	OS	50	0.1490	0.1625	0.1903	60.4	62.3	66.4	63.0	3.1
Fibro Pos	Control	20	0.0000	0.0000	0.0000	0.0	0.0	0.0	0.0	0.0
Fibro Pos	OS	50	0.1370	0.1554	0.1225	58.6	61.3	56.5	58.8	2.4

The fibroblast bound cell fraction of light density bone marrow cells represented 3–9 % of the starting nucleated cell population and adhered to polystyrene. Under standard MSC culture conditions, the fibroblast bound cell fraction yielded an MSC population as defined by flow cytometry, biological assays and morphology.

#### Example 6

##### MSC Isolation using Fibroblast and CD14 Cell Selection

10 Anti-fibroblast microbeads were from Miltenyi (Lot # NE6836). VS Column was from Miltenyi (Lot # 0231). Pre-separation filters 30  $\mu$ m were from Miltenyi (Lot # 55). Miltenyi buffer: phosphate buffered saline pH 7.2 supplemented with 0.5% BSA and 2mM EDTA. Anti-CD14 magnetic beads were from Dynal (Lot # A93900). Flow staining was performed following the manufacturer's suggested instructions. Flow analysis was performed using either  
15 the FACS Calibur or the FACS Vantage. Cell viability and cell number were determined using trypan blue.

Light density cells were isolated from a human bone marrow aspirate (donor #391) using Dendreon solution BDS72 following density centrifugation of  
20 the bone marrow aspirates. Aliquots were removed as controls for cell culture. The remaining cells were diluted to a cell count of  $2 \times 10^7$  cells/ml using Miltenyi buffer. Cells were incubated with IgG at 40  $\mu$ l per ml of cell suspension for 10 minutes at 4°C with mixing. Cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (80  $\mu$ l of buffer per  $10^7$  cells).  
25 Anti-fibroblast microbeads were added (20  $\mu$ l of beads per  $10^7$  cells). The mixture was incubated at 4°C for 15 minutes with mixing. Miltenyi buffer was added to dilute the mixture and wash the cells. The cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (1 ml of buffer per  $10^8$  cells). The VS column was primed following the manufacturer's instructions.  
30 The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'fibroblast unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this

effluent was added to the 'fibroblast unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'fibroblast bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger used to push the cells into the tube. A fresh column was primed and the cells in the 'fibroblast

5 bound' tube were added to the second column. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'fibroblast unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'fibroblast unbound'

10 fraction. The column was removed from the magnet and placed over a tube labeled 'fibroblast bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger used to push the cells into the tube. Cell counts and viability assays were performed. Unbound cells were placed in culture. Bound cells were incubated with Dynal Anti-CD14 magnetic beads at a cell concentration of  $2 \times 10^7$

15 cells per ml and a bead concentration of  $2 \times 10^7$  beads per ml for 1 hour at  $4^\circ\text{C}$  with mixing. The cell/bead suspension was placed next to the Dynal hand magnet for 2 minutes. After 2 minutes, the unattached cells were decanted into a tube labeled 'CD14 unbound'. The cell/bead tube was removed from the magnet and 5 ml of buffer added to resuspend the cells. The cell/bead suspension was placed next to

20 the Dynal hand magnet for 2 minutes. After 2 minutes, the unattached cells were decanted into a tube labeled 'CD14 unbound'. The cell/bead tube was removed from the magnet and 5 ml of buffer added to resuspend the cells. The cell/bead suspension was placed next to the Dynal hand magnet for 2 minutes. After 2 minutes, the unattached cells were decanted into a tube labeled 'CD14 unbound'.

25 The cell/bead tube was removed from the magnet and 5 ml of buffer added to resuspend the cells. The cells that remained attached to beads were counted and placed into culture in two  $10\text{ cm}^2$  wells. The cells in the unbound tube were placed next to the hand magnet for 2 minutes. Unattached cells were decanted, counted and plated into one  $10\text{ cm}^2$  well. Cells were harvested after 14 days. Cells

30 cultured from the light-density cells and the Fibroblast bound/CD14 bound fractions were stained for flow analysis.

Table 35

**Fibroblast Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$3.35 \times 10^8$	100
Fibroblast bound	$1.2 \times 10^7$	3.4
Fibroblast unbound	$3.51 \times 10^8$	105

Table 36

**CD14 Cell selection yields**

Fraction	Number of cells	% Yield
Fibroblast bound	$9 \times 10^6$	100
CD14 bound	$3.4 \times 10^6$	38
CD14 unbound	$1.9 \times 10^6$	21

Table 37

**Microscopic Examination of P0 Cultures on Day 14**

Fraction	Appearance
Light-density cells	100% confluent, phase bright cells seen
Fibroblast unbound	Round cells as singles and clumps
Fibroblast bound/CD14 bound	100% confluent with beads attached to some spindle shaped cells
Fibroblast bound/CD14 unbound	Round cells in singles and clumps

Table 38

**P0 Cell Harvester**

Fraction	Cells seeded/cm <sup>2</sup>	Cells harvested/cm <sup>2</sup>	% Yield
Light-density cells	$1.6 \times 10^5$	$7.6 \times 10^4$	47
Fibroblast unbound	$1.6 \times 10^5$	$1.6 \times 10^2$	0.1
Fibroblast bound/CD14 bound	$1.7 \times 10^5$	$4.0 \times 10^4$	23
Fibroblast bound/CD14 unbound	$1.9 \times 10^5$	$1.0 \times 10^3$	0.5

Table 39

**Flow analysis of P0 Cultures**

Fraction	% MSC phenotype	% CD45+ cells
Light-density cells	98.1	1.0
Fibroblast bound/CD14 bound	98.3	0.3

In order to further define the phenotype of the MSC precursor, a sequential cell selection was performed using Miltenyi microbeads to select fibroblast bound cells, followed by the use of Dynal magnetic beads to isolate CD14 bound cells from the fibroblast bound fraction. This is possible since the size of the microbead is too small to interfere with the Dynal selection.

Using this technique, the fibroblast bound/CD14 bound cell fraction was approximately 1.3% of the starting light-density cell population. The fibroblast bound/ CD14 bound cells, when placed in standard MSC culture conditions, adhered to polystyrene flasks and after 14 days of culture, yielded a MSC population as defined by flow analysis and morphology.

Based on this single selection, it would appear that the MSC precursor is a fibroblast+, CD14+ cell. This is unexpected since the culture expanded MSC is a fibroblast+, CD14 negative cell.

#### Example 7

##### MSC Isolation using Fibroblast Cell Selection and Flow Sorting

Anti-fibroblast microbeads were from Miltenyi (Lot # NE7105). VS Column was from Miltenyi (Lot # 0231). Pre-separation filters 30 um were from Miltenyi (Lot # 55). Miltenyi buffer: phosphate buffered saline pH 7.2 supplemented with 0.5% BSA and 2mM EDTA. Flow staining was performed following the manufacturer's suggested instructions. Flow analysis was performed using either the FACS Calibur or the FACS Vantage. Flow sorting was performed on the FACS Vantage. Determination of cell viability and cell number was done using trypan blue). Osteogenic potential was measured using the method described in Example 5 and with an *in vivo* osteogenic cube assay as described for example in U.S. Pat. No. 5,486,359. Adipogenic assays were performed according to methods described in Example 5.

The chondrogenesis assays was performed as follows. Cells were pelleted in a 15ml conical tube ( $2.5 \times 10^5$  cells/pellet) in chondrogenic media, consisting of high glucose, dexamethasone and TGF- $\beta$ 3. The pellets were submitted to histology for embedding, thin sectioning and histochemical staining. The presence

of chondrocytes was detected using Toluidine Blue, which stains for proteoglycans and with an antibody specific for type II collagen.

Light density cells were isolated from human bone marrow aspirate (donors # 401 and # 438) using Dendreon solution BDS72 following density centrifugation of the bone marrow aspirates. Aliquots were removed as controls for flow analysis and cell culture. The remaining cells were diluted to a cell count of  $2 \times 10^7$  cells/ml using Miltenyi buffer. Cells were incubated with IgG at 40 ul per ml of cell suspension for 10 minutes at 4°C with mixing. Cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (80 ul of buffer per  $10^7$  cells). Anti-fibroblast microbeads were added (20 ul of beads per  $10^7$  cells). The mixture was incubated at room temperature for 30 minutes with mixing. Miltenyi buffer was added to dilute the mixture and wash the cells. The cells were centrifuged for 10 minutes at 1000 RPM. The cell pellet was resuspended in Miltenyi buffer (1 ml of buffer per  $10^8$  cells). The VS column was primed following the manufacturer's instructions. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'fibroblast unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'fibroblast unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'fibroblast bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger was used to push the cells into the tube. A fresh column was primed and the cells in the 'fibroblast bound' tube was added to the second column. The pre-filter was primed following the manufacturer's instructions. The cell/microbead suspension was added to the pre-filter and the suspension drained through the column. The effluent was collected as the 'fibroblast unbound' fraction. The column was rinsed twice with 3 ml of Miltenyi buffer and this effluent was added to the 'fibroblast unbound' fraction. The column was removed from the magnet and placed over a tube labeled 'fibroblast bound'. Five ml of Miltenyi buffer was added to the column and a syringe plunger was used to push the cells into the tube. Cell counts and viability assays were performed.

Cells from the fibroblast unbound fraction were stained for flow analysis. Cells from the fibroblast bound fraction were incubated with anti-SH3, anti-CD45

and anti-CD14. Cells were sorted: Donor 401 was sorted into two groups: population 1 and population 2. Donor 438 was sorted into three groups: population 1, population 1a and population 2. Cells from the fibroblast unbound fraction (Donor 401 only) and the sorted populations were placed in culture. P0 cells were harvested and passaged. P1 cells from the light-density and population 1 cultures on donor # 401 were stained for flow cytometry and placed in the *in vivo* cube assay. P1 cells for donor # 438 were passaged. P2 cells from the light density, population 1 and population 1a cultures on donor # 438 were stained for flow cytometry and placed in the *in vitro* biological assays (osteogenic, adipogenic and chondrogenic).

Table 40

**Donor 401 Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$6.6 \times 10^8$	100
Fibroblast bound	$2.9 \times 10^7$	4.4
Fibroblast unbound	$5.4 \times 10^8$	82

Table 41

**Donor 438 Cell selection yields**

Fraction	Number of cells	% Yield
Light-density cells	$8.5 \times 10^8$	100
Fibroblast bound	$2.0 \times 10^7$	2.4
Fibroblast unbound	$7.9 \times 10^8$	93

Table 42

**Fibroblast Bound Cells after Flow Sorting**

Fraction	Surface Phenotype Flow Profile	Number of Cells
Donor 401 Population 1	SH3+ bright; CD14 dim	$1.1 \times 10^4$
Donor 401 Population 2	SH3+ bright; CD14 bright	$2.0 \times 10^4$
Donor 438 Population 1	SH3+ bright; CD14 dim	$1.5 \times 10^4$
Donor 438 Population 1a	SH3+ bright; CD45 dim	$1.5 \times 10^4$
Donor 438 Population 2	SH3+ bright; CD14 bright	$4.8 \times 10^5$



Table 43

**Donor 401: Microscopic Examination of P0 Cultures on Day 12**

<b>Fraction</b>	<b>Appearance</b>
Light-density cells	60% confluent, phase bright cells seen
Fibroblast unbound	Round cells with rare spindle shaped cells seen
Fibroblast bound Population 1	100% confluent, phase bright cells seen
Fibroblast bound Population 2	Round cells with rare spindle shaped cells seen

5

Table 44

**Donor 438: Microscopic Examination of P0 Cultures on Day 14**

<b>Fraction</b>	<b>Appearance</b>
Light-density cells	90 % confluent, phase bright cells seen
Fibroblast bound Population 1	100 % confluent, phase bright cells seen
Fibroblast bound Population 1a	100% confluent, phase bright cells seen
Fibroblast bound Population 2	Debris

Table 45

**Donor 401: P0 (Day 12 cultures) Cell Harvests**

<b>Fraction</b>	<b>Cells Seeded/cm<sup>2</sup></b>	<b>Cells Harvested/cm<sup>2</sup></b>	<b>% Yield</b>
Light-density cells	$8.0 \times 10^4$	$1.7 \times 10^4$	21
Fibroblast unbound	$8.0 \times 10^4$	$6.7 \times 10^2$	0.08
Population 1	$8.9 \times 10^2$	$1.7 \times 10^3$	196
Population 2	$1.7 \times 10^3$	$7.1 \times 10^2$	42

10

Table 46

**Donor 438: P0 (Day 14 cultures) Cell Harvests**

<b>Fraction</b>	<b>Cells Seeded/cm<sup>2</sup></b>	<b>Cells Harvested/cm<sup>2</sup></b>	<b>Fold Expansion</b>
Light-density cells	$8.0 \times 10^4$	$4.5 \times 10^4$	0.6
Population 1	$6.0 \times 10^2$	$4.2 \times 10^4$	70
Population 1a	$6.0 \times 10^2$	$4.6 \times 10^4$	77
Population 2	$9.6 \times 10^3$	0	0

Table 47

**P1 (Day 7 cultures) Cell Harvests**

Fraction	Cells Seeded/cm <sup>2</sup>	Cells Harvested/cm <sup>2</sup>	Fold Expansion
401 Light-density cells	$6.7 \times 10^3$	$3.6 \times 10^4$	5
401 Population 1	$0.3 \times 10^3$	$2.5 \times 10^4$	90.0
438 Light-density cells	$6.1 \times 10^3$	$2.3 \times 10^4$	4
438 Population 1	$5.7 \times 10^3$	$2.1 \times 10^4$	4
438 Population 1a	$6.2 \times 10^3$	$2.6 \times 10^4$	5

Table 48

**Flow analysis of P1 Cultures Donor 401(1)**

Fraction	% MSC phenotype	% CD45+
Light-density cells	99.2	0.2
Population 1	99.8	<0.1

5

Table 49

**In vivo Cube Assay Results Donor 401(1)**

Fraction	Average Score for 3 cubes
Light-density cells	0.3
Population 1	0.5
Negative Control	0

Table 50

**Donor 438: P2 (Day 7 cultures) Cell Harvest**

Fraction	Cells seeded/cm <sup>2</sup>	Cells Harvests/cm <sup>2</sup>	Fold Expansion
Light-density cells	$5.4 \times 10^3$	$2.1 \times 10^4$	3.8
Population 1	$5.3 \times 10^3$	$2.4 \times 10^4$	4.5
Population 1a	$5.3 \times 10^3$	$2.2 \times 10^4$	4.2

10

Table 51

**Flow analysis of P2 Cultures Donor 438(2)**

Fracti n	% MSC phen type	% CD45+
Light-density cells	99.9	< 0.1
Population 1	99.9	< 0.1
Population 1a	99.9	< 0.1

Table 52

**Donor 438(2): Adipogenesis Results**

Fraction	% Area with Vacuoles		
	Adipogenic Medium	Adipogenic Maintenance Medium	Human MSC Medium
Light-density cells	40	0	0
Population 1	40	0	0
Population 1a	40	0	0

All samples measured positive as evaluated by established acceptance criteria.

5

Table 53

**Donor 438(2): Osteogenesis Results**

Fraction	Calcium ug/dish		Calcium ug/million cells	
	Control	OS	Control	OS
Light-density cells	1.0	16.2	5.1	12.9
Population 1	1.1	18.0	7.2	15.1
Population 1a	1.1	14.4	5.8	12.8

All samples measured positive as evaluated by established acceptance criteria.

Table 54a

**Donor 438(2): Chondrogenesis Results; Toluidine Blue Results**

Fraction	Toluidine Blue (%)			
	Day 7	Day 14	Day 21	Day 28
Light-density cells	60	40	70	100
Population 1	70	100	90	100
Population 1a	90	100	100	100

10

Table 54b

**Donor 438(2): Chondrogenesis Results; Collagen Type II Results**

Fracti n	Anti-Collagen Type II (%)			
	Day 7	Day 14	Day 21	Day 28
Light-density cells	0	0	30	50
Population 1	0	100	70	80
Population 1a	0	90	100	80

Table 54c

**Donor 438(2): Chondrogenesis Results; Pellet Size**

Fraction	Diameter (millimeters)			
	Day 7	Day 14	Day 21	Day 28
Light-density cells	1.0	0.8	0.9	1.0
Population 1	0.8	1.2	1.5	1.5
Population 1a	1.0	1.4	1.6	1.5

All samples measured positive as evaluated by established acceptance criteria.

5

To further define the phenotype of the MSC precursor, fibroblast bound light-density bone marrow cells were stained with anti-SH3 and anti-CD14 and sorted using flow cytometry.

This resulted in the sorting of population 1, population 1a and population 2.  
 10 All three of these populations were SH3+ bright. Population 1 was CD14+ with a dim staining intensity. Population 1a was CD45+ with a dim staining intensity. Population 2 was CD14+ with a bright staining intensity. MSCs were cultured from Population 1 and Population 1a cell fractions, as confirmed from flow analysis and *in vitro* and *in vivo* biological assays. It was estimated that one  
 15 Population 1/1a cell is present in  $10^5$  light-density bone marrow cells.

## CLAIMS

1. An isolated population of human mesenchymal stem cells comprising human mesenchymal stem cells which are CD45+.
- 5 2. The isolated population of human mesenchymal stem cells of claim 1 wherein the human mesenchymal stem cells which are CD45 + are further reactive with at least one monoclonal antibody selected from the group consisting of SH2 antibody, SH3 antibody and SH4 antibody.
- 10 3. The isolated cell population of claim 2 wherein the human mesenchymal stem cells are SH3+and CD45+.
4. The isolated cell population of claim 2 wherein the human mesenchymal stem cells are SH2+ and CD45+.
- 15 5. A composition comprising an isolated population of human mesenchymal stem cells which bear a fibroblast surface marker.
6. The composition of claim 5 wherein the human mesenchymal stem cells are  
20 also CD45+.
7. The composition of claim 6 further comprising CD14+ cells.
8. A process of isolating a population of human mesenchymal stem cells which  
25 are CD45+, comprising
  - (a) obtaining an enriched population of human mesenchymal stem cells; and
  - (b) selecting from the cell population of (a) cells that are CD45+.
9. A process of isolating a population of human mesenchymal stem cells which  
30 are SH3+ and CD45+, comprising
  - (a) obtaining an enriched population of human mesenchymal stem cells that are SH3+; and
  - (b) selecting from the cell population of (a) cells that are CD45+.

10. A process of isolating a population of human mesenchymal stem cells which are SH2+ and CD45+, comprising
- (a) obtaining an enriched population of human mesenchymal stem cells that are SH2+; and
  - 5 (b) selecting from the cell population of (a) cells that are CD45+.
11. A process of isolating a population of human mesenchymal stem cells which are fibroblast+, comprising
- (a) obtaining an enriched population of human mesenchymal stem
  - 10 cells; and
  - (b) selecting from the cell population of (a) cells that are fibroblast+.
12. The method of claim 11, further comprising selecting from the fibroblast+ cell population cells that are SH3+.
- 15 13. The method of claim 12, further comprising selecting from the fibroblast+/SH3+ cell population cells that are CD45+.
- 20 14. A method for treating a patient, comprising administering human mesenchymal stem cells which are CD45+ to a patient in need thereof.
15. The method of claim 14 wherein the human mesenchymal stem cells are SH3+ and CD45+.
- 25 16. The method of claim 14 wherein the human mesenchymal stem cells are SH2+ and CD45+.
17. The method of claim 14 wherein the mesenchymal stem cells are
- 30 administered to generate bone formation.
18. The method of claim 14 wherein the mesenchymal stem cells are administered to treat or repair a connective tissue defect in the patient.

19. The method of claim 18 wherein the defect is a bone defect.
20. The method of claim 18 wherein the defect is a cartilage defect.
- 5 21. The method of claim 14 wherein the mesenchymal stem cells are administered to enhance hematopoietic stem or progenitor cell engraftment in an individual in need thereof.
22. A method for treating a patient, comprising administering human  
10 mesenchymal stem cells which are fibroblast+ to a patient in need thereof.
23. Isolated CD45+ human mesenchymal stem cells transfected with exogenous genetic material encoding a protein to be expressed.
- 15 24. Isolated fibroblast+ human mesenchymal stem cells transfected with exogenous genetic material encoding a protein to be expressed.

FIG.1A  
Pre-fractionation

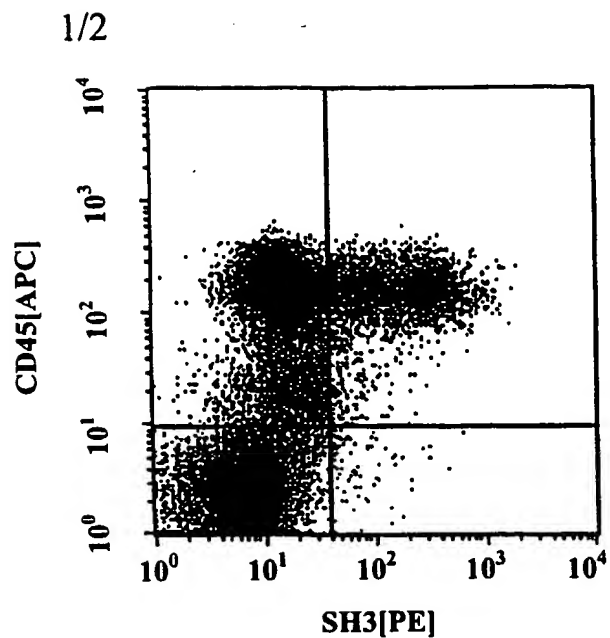


FIG.1B  
SH3 negative fraction

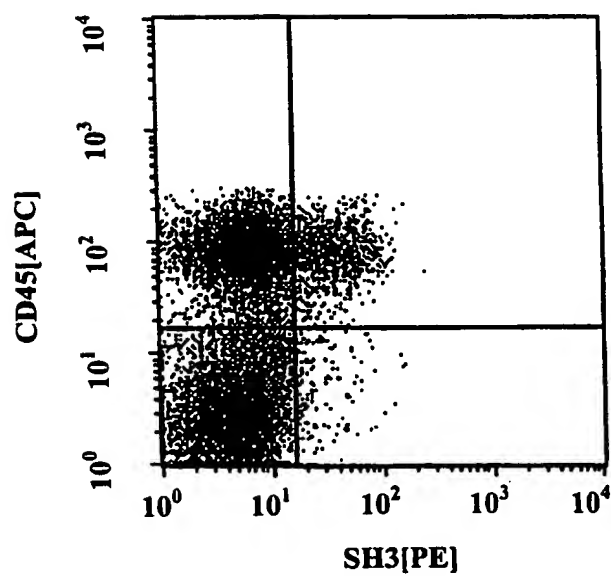
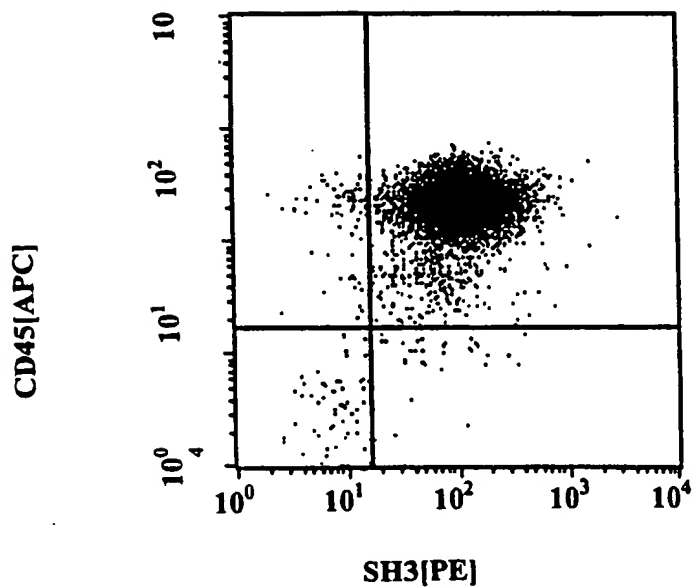


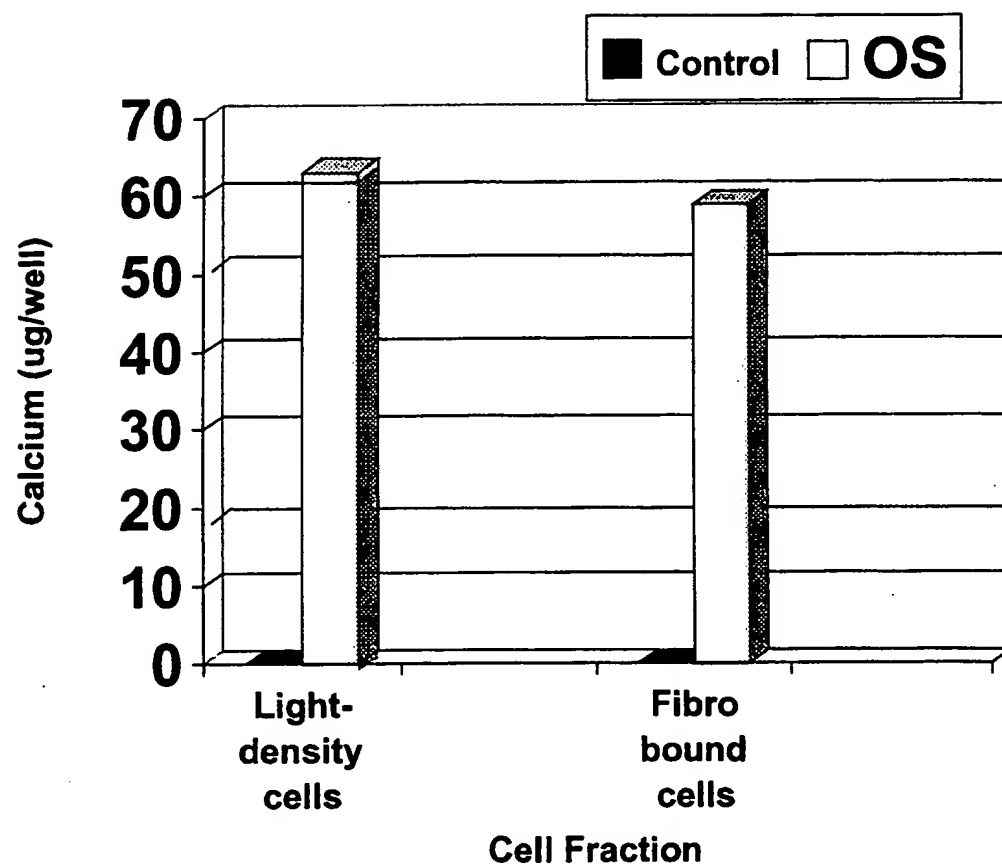
FIG.1C  
SH3 positive fraction





2/2

FIG. 2



# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/11975

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N5/06 A61K35/28 //C07K14/705

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 96 38482 A (OSIRIS THERAPEUTICS INC) 5 December 1996 (1996-12-05) page 13, paragraph 3 - page 17, paragraph 1	1-7
X	US 5 654 186 A (CERAMI ANTHONY ET AL) 5 August 1997 (1997-08-05) column 2, line 45 - column 3, line 20	1-24



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

### \* Special categories of cited documents :

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# INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/US 99/11975

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